

# **Modern Biotechnology in the Clinical Microbiology Lab**

## **Faster Methods for Getting Actionable Intelligence to Physicians**

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# Clinical Scenario #1

50 year old female, recently Army retiree presents 1 year out from a right total knee replacement with worsening knee pain and mild swelling along with some night sweats. She is taken to the OR and the hardware is removed and sent down to the lab for culture...

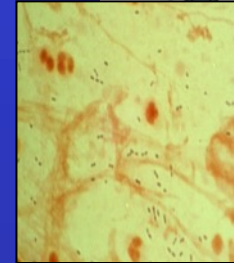
# Bacterial Culture and Identification Timeline



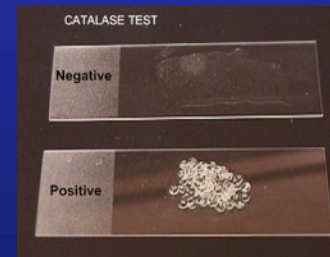
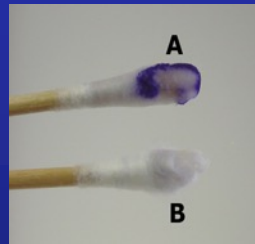
1 hr



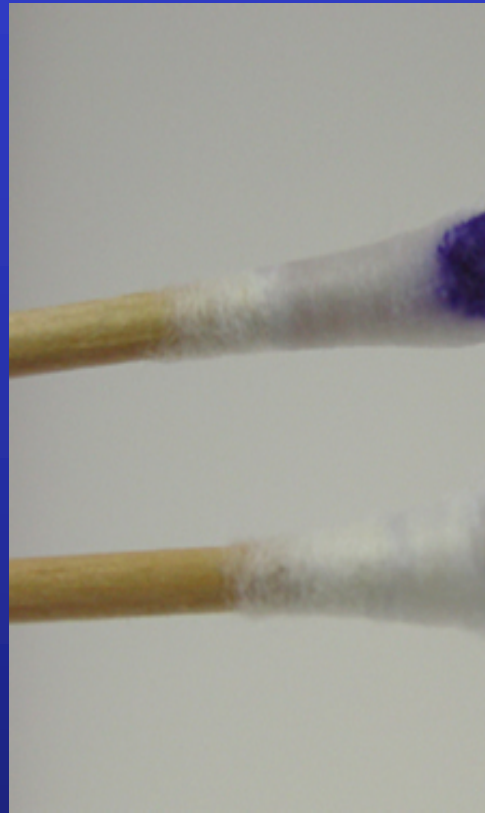
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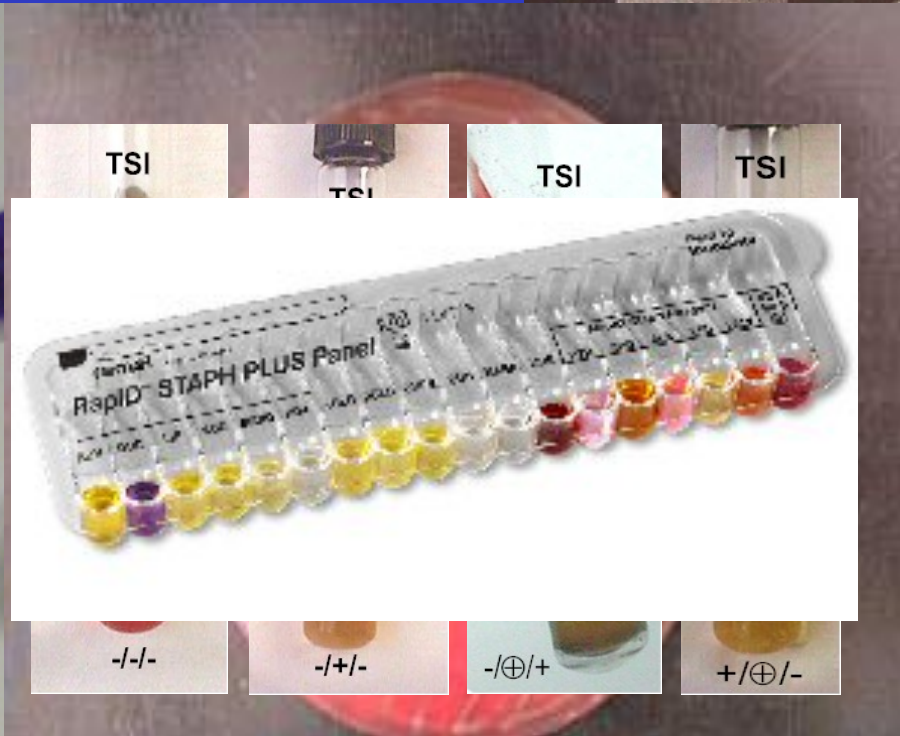
~18 hrs



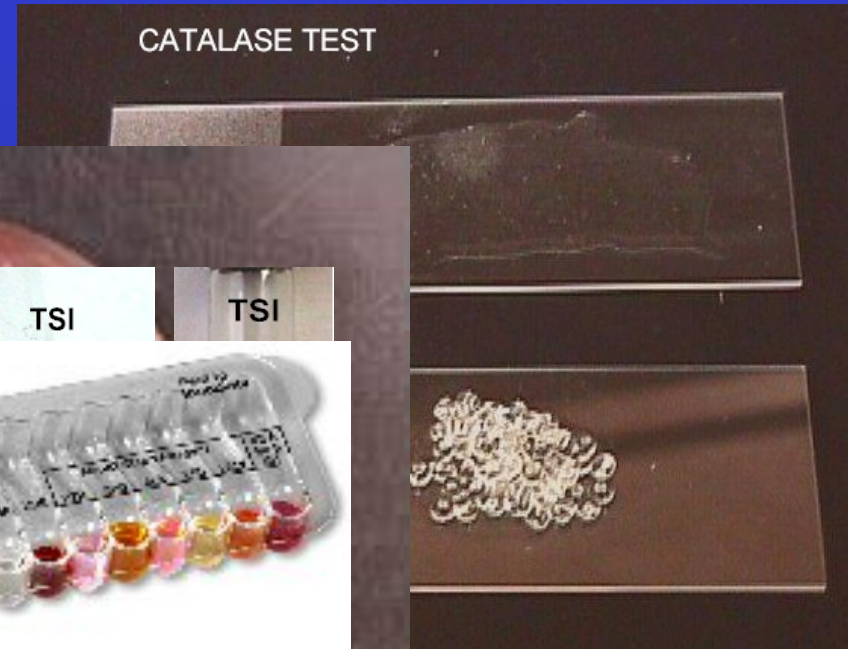
# Biochemically-Based Identification Methods



Indole test (*E.coli*)



Lancefield Group typing (*Strep species*)



Test (Staph/Strep)



# Automated Biochemical Systems



- Identification and Susceptibilities
- ID based on biochemical profile
- Antibiotic breakpoints based on CLSI standards
- Some susceptibilities reported out as standard, some withheld and reported out by exception

## Examples:

- **Biomerieux Vitek**
- **Becton Dickinson Pheonix**
- **Beckman Coulter MicroScan**

- Pure culture required
- ~18 hour cycle

# Laboratory Diagnosis

- Biochemical Tests

- Triple Sugar Iron Agar / Kligler Iron Agar
- Indole production
- Methyl Red test
- Voges-Proskauer test
- Citrate utilization
- Nitrate reduction
- Urease production
- Oxidase activity
- Carbohydrate fermentation (Adonitol, Arabinose, Inositol, Sucrose)
- Decarboxylation of Lysine, Ornithine, and Arginine (amino acids)
- Phenylalanine Deaminase production
- o-Nitrophenyl- $\beta$ -D-galctopyranoside (ONPG)
  - Tests for  $\beta$ -galactosidase; helpful in identifying late lactose fermenters
- Hydrogen Sulfide production
- Motility

**Make up the 13-tube  
biochemical ID panel**

# Biochemical Testing

	LF?	Motility	Indole	Methyl Red	Voges Proskauer	H <sub>2</sub> S	Citrate	Urease
<i>K. pneumoniae</i>	Yes	-	-	-	+	-	+	+
<i>K. oxytoca</i>	Yes	-	+	-	+	-	+	+
<i>Y. pestis</i>	No	-	-	+	-	-	-	-
<i>Y. enterocolitica</i>	No	+ (25 C)	V(50%)	+	-	-	-	+
<i>E. coli</i>	Yes	+	+	+	-	-	-	-
<i>Shigella</i>	No	-	-/+	+	-	-	-	-
<i>Salmonella</i>	No	+	-	+	-	+	+	-
S. Typhi	No	+	-	+	-	+ (wk)	-	-
P. mirabilis	No	+ (sw.)	-	+	-	+	+/-	++
P. vulgaris	No	+ (sw.)	+	+	-	+	-	++



AsheX © 2004-2018 Non-Fermenter ID Matrix:	Motility	Oxidase	Catalase	Yellow Pig	Pink Pig	Beta Hem	Growth on	DNase	Starch	Lecithinas	Lipase	PYR	LAP	ESC Spot
<i>Achromobacter denitrificans</i>	99.00	99.00	99.00	1.00	1.00	16.67	99.00	1.00	1.00	1.00	1.00	55.56	99.00	1.00
<i>Achromobacter piechaudii</i>	78.57	99.00	99.00	1.00	1.00	14.29	99.00	1.00	1.00	1.00	1.00	50.00	99.00	1.00
<i>Achromobacter xylosoxidans</i>	87.50	99.00	99.00	1.00	1.00	1.56	99.00	1.00	1.00	1.00	1.00	78.72	99.00	1.00
<i>Acidovorax temperans</i>	99.00	99.00	99.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00	99.00	99.00	1.00
<i>Acinetobacter baumannii</i> complex	1.00	1.00	99.00	1.00	1.00	1.00	99.00	1.00	1.00	1.00	27.27	10.00	99.00	1.00
<i>Acinetobacter haemolyticus</i>	1.00	1.00	99.00	1.00	1.00	99.00	99.00	1.00	1.00	1.00	99.00	1.00	99.00	1.00
<i>Acinetobacter lwoffii</i>	1.00	1.00	99.00	1.00	1.00	1.00	82.61	1.00	1.00	1.00	1.00	22.22	99.00	1.00
<i>Acinetobacter species saccharolytic</i>	50.00	1.00	99.00	1.00	1.00	99.00	99.00	1.00	1.00	1.00	99.00	50.00	50.00	50.00
<i>Alcaligenes faecalis</i>	99.00	99.00	99.00	1.00	1.00	1.00	99.00	1.00	1.00	1.00	1.00	1.00	99.00	1.00
<i>Bergeyella zoohelcum</i>	1.00	83.33	50.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00	99.00	1.00
<i>Bordetella avium</i>	99.00	99.00	99.00	1.00	1.00	1.00	99.00	1.00	1.00	1.00	1.00	99.00	99.00	1.00
<i>Bordetella bronchiseptica</i>	84.21	99.00	99.00	1.00	1.00	1.00	99.00	1.00	1.00	1.00	1.00	22.22	99.00	1.00
<i>Bordetella hinzii</i>	99.00	99.00	50.00	50.00	1.00	1.00	99.00	1.00	1.00	1.00	1.00	1.00	99.00	1.00
<i>Bordetella holmesii</i>	1.00	12.50	37.50	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00	87.50	1.00
<i>Bordetella parapertussis</i>	1.00	1.00	99.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00	99.00	1.00
<i>Bordetella trematum</i>	99.00	60.00	99.00	1.00	1.00	1.00	99.00	1.00	1.00	1.00	1.00	20.00	99.00	1.00
<i>Brevundimonas diminuta</i>	99.00	99.00	92.86	1.00	1.00	1.00	92.86	1.00	1.00	1.00	1.00	1.00	75.00	1.00
<i>Brevundimonas vesicularis</i>	99.00	93.33	60.00	33.33	1.00	8.33	46.67	6.67	66.67	1.00	1.00	28.57	85.71	42.86
<i>Pandoraea apista</i>	99.00	1.00	99.00	1.00	1.00	1.00	99.00	1.00	1.00	1.00	1.00	1.00	99.00	1.00
<i>Pseudomonas aeruginosa</i>	67.92	98.11	99.00	1.00	1.89	73.91	99.00	9.43	24.53	10.87	30.43	34.21	97.37	1.00
<i>Pseudomonas alcaligenes</i>	99.00	99.00	99.00	1.00	1.00	1.00	88.89	1.00	1.00	1.00	1.00	99.00	99.00	11.11
<i>Pseudomonas fluorescens</i>	90.91	95.45	99.00	4.55	9.09	72.22	99.00	4.55	50.00	70.00	36.84	23.08	99.00	1.00
<i>Pseudomonas luteola</i>	99.00	1.00	92.31	84.62	1.00	1.00	92.31	15.38	53.85	1.00	1.00	1.00	99.00	99.00
<i>Pseudomonas mendocina</i>	99.00	99.00	75.00	25.00	8.33	1.00	99.00	1.00	16.67	1.00	33.33	1.00	99.00	1.00
<i>Pseudomonas oryzae</i> habitans	99.00	1.00	99.00	88.24	1.00	8.33	99.00	1.00	64.71	1.00	1.00	36.36	99.00	1.00
<i>Pseudomonas pseudoalcaligenes</i>	69.23	99.00	84.62	1.00	1.00	1.00	99.00	1.00	7.69	1.00	1.00	1.00	99.00	1.00
<i>Pseudomonas putida</i>	99.00	96.88	96.88	18.75	1.00	15.38	99.00	1.00	25.00	3.70	3.70	1.00	99.00	1.00
<i>Pseudomonas stutzeri</i>	91.30	99.00	95.65	65.22	1.00	1.00	99.00	1.00	86.96	1.00	83.33	6.25	93.75	1.00
<i>Pseudomonas stutzeri</i> (Vb-3)	99.00	99.00	99.00	33.33	16.67	1.00	99.00	1.00	99.00	1.00	99.00	50.00	50.00	50.00
<i>Psychrobacter immobilis</i> (asaccharolytic)	1.00	99.00	99.00	1.00	1.00	20.00	80.00	1.00	1.00	1.00	40.00	1.00	80.00	1.00
<i>Psychrobacter immobilis</i> (saccharolytic)	1.00	99.00	99.00	1.00	1.00	20.00	99.00	1.00	1.00	1.00	60.00	1.00	99.00	1.00
<i>Psychrobacter phenylpyruvicus</i>	1.00	99.00	99.00	1.00	1.00	1.00	80.00	1.00	1.00	1.00	33.33	1.00	99.00	1.00
<i>Ralstonia mannitolilytica</i>	99.00	99.00	14.29	1.00	1.00	1.00	71.43	1.00	14.29	1.00	42.86	28.57	99.00	1.00
<i>Ralstonia pickettii</i> (Va-1)	78.57	99.00	1.00	14.29	1.00	1.00	57.14	1.00	35.71	1.00	70.00	55.56	99.00	10.00
<i>Ralstonia pickettii</i> (Va-2)	99.00	99.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00	60.00	20.00	99.00	1.00
<i>Rhizobium</i> ( <i>Agrobacterium</i> ) <i>radiobacter</i>	84.62	92.31	99.00	1.00	1.00	33.33	72.73	1.00	1.00	1.00	1.00	99.00	83.33	33.33
<i>Roseomonas species</i>	15.38	61.54	99.00	1.00	99.00	1.00	7.69	1.00	69.23	1.00	1.00	14.29	57.14	1.00
<i>Shewanella algae</i>	99.00	83.33	99.00	1.00	16.67	25.00	99.00	99.00	1.00	1.00	75.00	66.67	99.00	1.00
<i>Shewanella putrefaciens</i>	80.00	99.00	60.00	20.00	20.00	1.00	99.00	99.00	1.00	1.00	1.00	1.00	99.00	1.00
<i>Sphingobacterium multivorum</i>	50.00	99.00	99.00	33.33	1.00	20.00	99.00	16.67	83.33	1.00	60.00	80.00	99.00	40.00
<i>Sphingobacterium spiritivorum</i>	1.00	99.00	88.89	11.11	1.00	1.00	1.00	99.00	1.00	1.00	1.00	1.00	99.00	99.00
<i>Sphingobacterium thalophilum</i>	1.00	99.00	99.00	1.00	1.00	50.00	99.00	99.00	99.00	50.00	50.00	50.00	50.00	50.00
<i>Sphingomonas paucimobilis</i>	50.00	55.00	85.00	99.00	1.00	41.67	5.00	25.00	75.00	1.00	1.00	1.00	99.00	99.00
<i>Stenotrophomonas maltophilia</i>	88.00	28.00	94.00	36.00	1.00	22.50	96.00	82.00	1.00	1.00	71.11	1.00	94.87	97.44



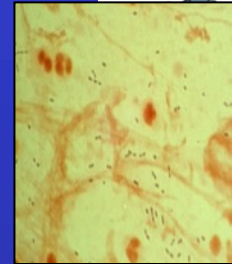
# Bacterial Culture and Identification Timeline



1 hr



1 hr

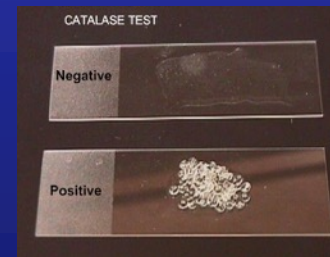
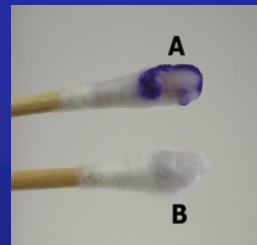


**Total time to ID: ~48 hours**  
**Total time to antibiotic profile: ~48 hours**

~18 hrs



~24 hrs



# Antibiotic Therapies

## Knowing the species matters

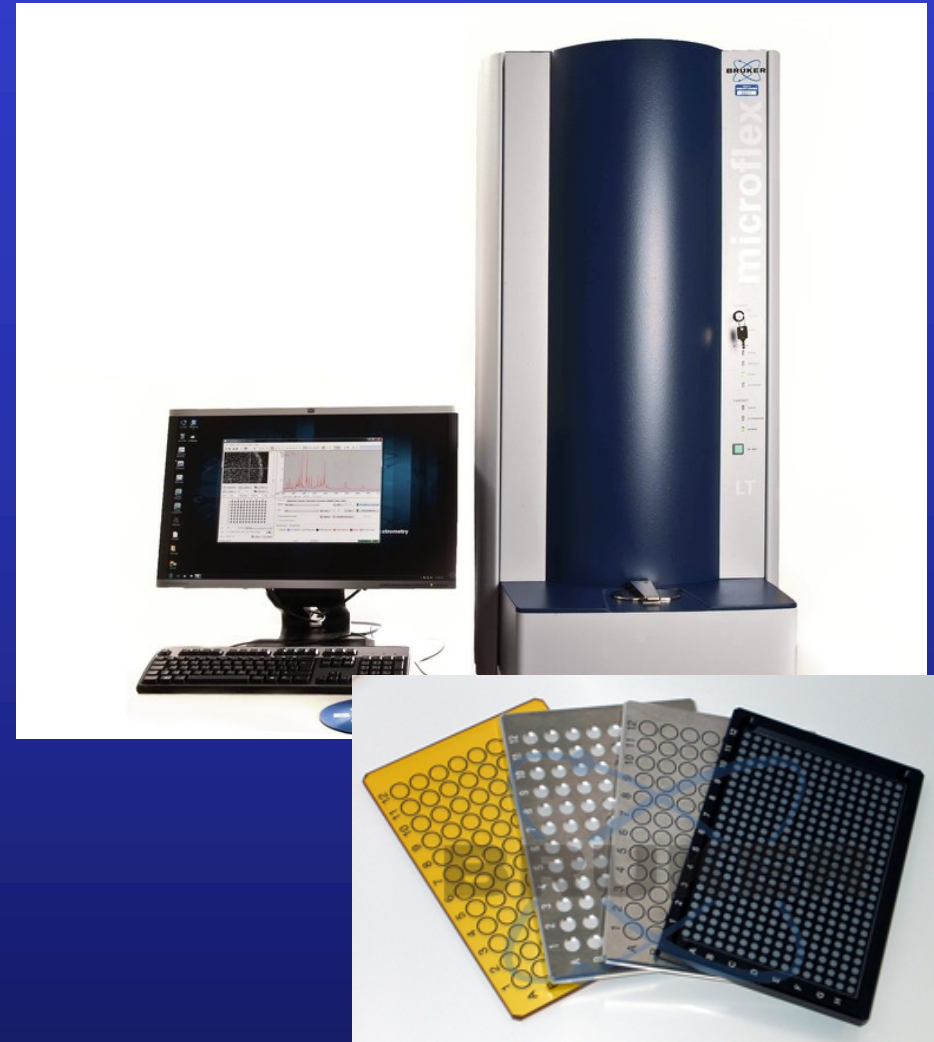
**Table 4**

Antimicrobial stewardship recommendations for the treatment of BSI caused by organisms identified by the FilmArray® BCID panel<sup>a</sup>.

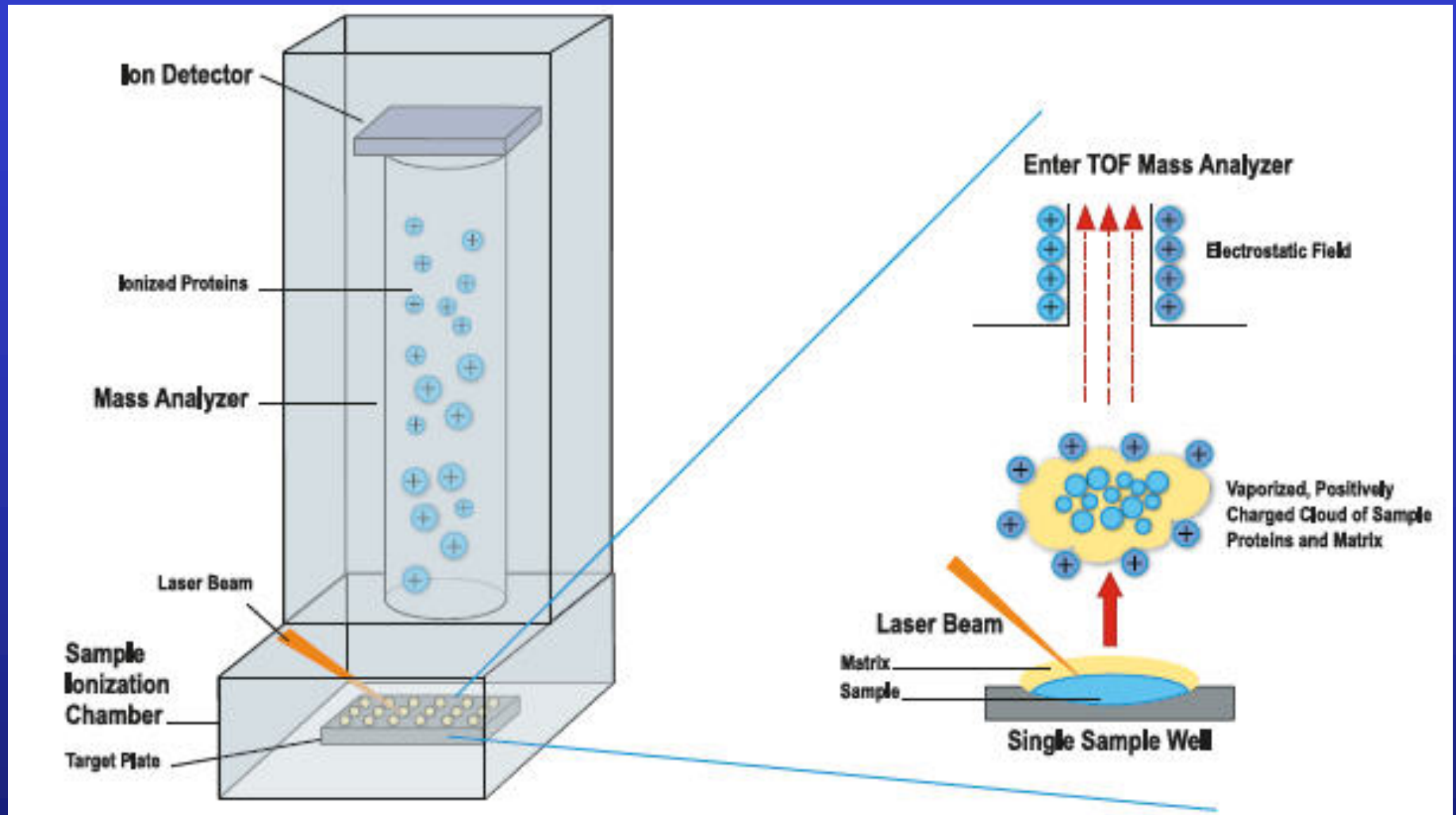
BCID result	No. positive BC sets <sup>b</sup>	Preferred therapy (alternative therapy)	Comments <sup>c</sup>
<i>Staphylococcus</i> ; <i>mecA</i> negative or positive	1 of multiple	None, likely contaminant	If severely ill, consider antibiotic therapy until more results
<i>Staphylococcus</i> ; <i>mecA</i> negative	2 or more	OXA 2 g q4h (CFZ 2 g q8h)	NA
<i>Staphylococcus</i> ; <i>mecA</i> positive	2 or more	VAN 15 mg/kg q12h (DAP 6 mg/kg q24h)	NA
<i>Staphylococcus aureus</i> ; <i>mecA</i> negative	1 or more	OXA 2 g q4h (CFZ 2 g q8h)	NA
<i>Staphylococcus aureus</i> ; <i>mecA</i> positive	1 or more	VAN 15 mg/kg q12h (DAP 6 mg/kg q24h)	NA
<i>Streptococcus</i>	1 of multiple	None, likely contaminant	If severely ill, consider antibiotic therapy until more results
<i>Streptococcus</i>	2 or more	CRO 2 g q24h	NA
<i>Streptococcus pyogenes</i> , <i>S. agalactiae</i>	1 or more	PEN 3 million units q4h (AMP 2 g IV q4h; CRO 2 g IV q24h)	Beta-hemolytic streptococci are routinely susceptible to PEN
<i>Streptococcus pneumoniae</i> (non-CNS)	1 or more	PEN 3 million units q4h (AMP 2 g IV q4h)	NA
<i>Streptococcus pneumoniae</i> (CNS)	1 or more	CRO 2 g q12h and VAN 15 mg/kg q12h	Continue VAN until susceptibilities are available
<i>Enterococcus</i> ; <i>vanA/B</i> negative	1 or more	VAN 15 mg/kg q12h	NA
<i>Enterococcus</i> ; <i>vanA/B</i> positive	1 or more	LZD 600 mg q12h (DAP 6–8 mg/kg q24h)	DAP is less active than LZD
<i>Listeria monocytogenes</i>	1 or more	AMP 2 g q4h	Consider SXT for individuals with beta-lactam allergies
Enterobacteriaceae (only)	1 or more	TZP 4.5 g q8h over 4 h (FEP 1 g q6h)	Consider stopping non-beta lactam if on combination therapy
<i>Escherichia coli</i>	1 or more	CRO 2 g q24h (ERT 1 g q24h; for severely ill)	CRO, 97% susceptible; ERT, 99% susceptible
<i>Klebsiella pneumoniae</i>	1 or more	CRO 2 g q24h	CRO, 98% susceptible
<i>Klebsiella oxytoca</i>	1 or more	ERT 1 g q24h (CRO and TZP)	ERT, 100% susceptible, CRO and TZP, 88% susceptible
<i>Serratia marcescens</i>	1 or more	CRO 2 g q24h (FEP 1 g q6h)	CRO, 96% susceptible; FEP, 99% susceptible
<i>Enterobacter cloacae</i> complex	1 or more	FEP 1 g q6h (ERT 1 g q24h; for severely ill)	FEP, 90% susceptible; ERT, 99% susceptible
<i>Proteus</i>	1 or more	CRO 2 g q24h	CRO, 98% susceptible
<i>Acinetobacter baumannii</i>	1 or more	MEM 500 mg q6h ± GEN 7 mg/kg daily	MEM, 92.5% susceptible, consider adding GEN for severely ill
<i>Pseudomonas aeruginosa</i>	1 or more	TZP 4.5 g q8h over 4 h ± TOB 7 mg/kg daily	TZP, 92.5% susceptible; consider adding TOB for severely ill
<i>Neisseria meningitidis</i>	1 or more	PEN 4 million units q4h (CRO 2 g q12h)	NA
<i>Haemophilus influenzae</i>	1 or more	SAM 3 g q6h (CRO 2 g q24h)	NA
<i>Candida albicans</i>	1 or more	FLC 800 mg load, 400 mg daily <sup>d</sup>	93% susceptible; 3% susceptible dose-dependent
<i>Candida parapsilosis</i>	1 or more	FLC 800 mg load, 400 mg daily <sup>d</sup>	91% susceptible; 6% susceptible dose-dependent
<i>Candida glabrata</i> , <i>C. krusei</i> , <i>C. tropicalis</i>	1 or more	MFG 100 mg q24h	99–100% susceptible
<i>mecA</i>	1 or more <sup>e</sup>	VAN 15 mg/kg q12h	Marker for methicillin-resistant <i>Staphylococcus</i>
<i>vanA/B</i>	1 or more <sup>f</sup>	LZD 600 mg q12h	Marker for VAN-resistant <i>Enterococcus</i>
<i>bla<sub>KPC</sub></i>	1 or more	Consult Infectious Disease Service; COL ± TGC	Marker for carbapenem-resistant Enterobacteriaceae

# Matrix Assisted Laser Desorption Ionization-Time Of Flight (MALDI-TOF)

- **Protein-based identification**
- **Accuracy comparable to Nucleic Acid Sequencing**
  - >98% accuracy compared to 16S RNA sequencing
- **Much faster than traditional methods**
  - ~200 IDs an hour from a pure colony (**requires initial culturing**)
- **Cost effective (with large initial capital investment)**
  - Less than a dollar per test
- **Two commercially-available, FDA-approved platforms**
  - Bruker MS Biotyper
  - Biomerieux Vitek MS



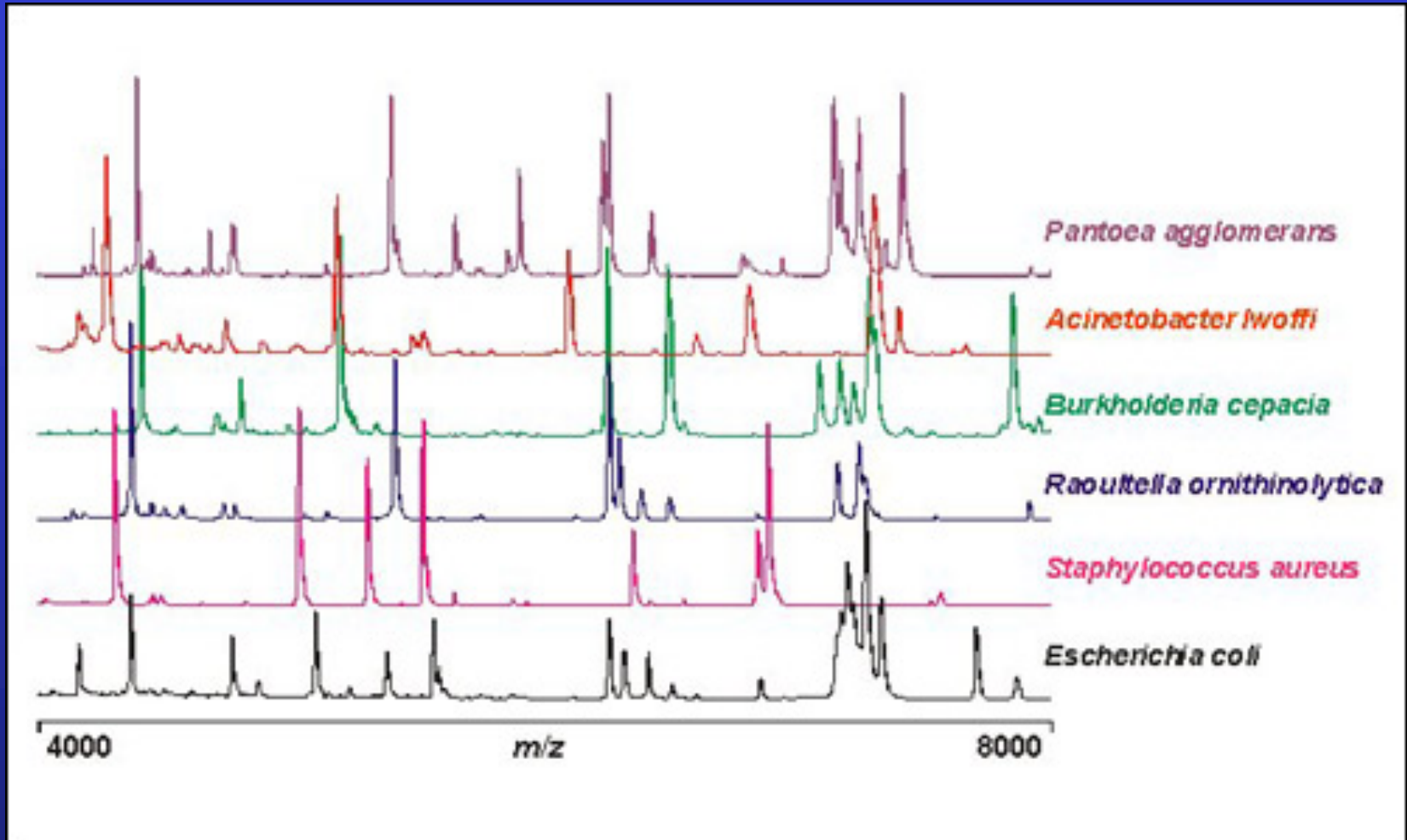
# MALDI-TOF



- Measures highly abundant proteins found in all organisms (ex. 16S Ribosomal proteins)
- Creates a spectra fingerprint to compare against a database of known organisms



# MALDI-TOF



- Measures highly abundant proteins found in all organisms (ex. 16S Ribosomal proteins)
- Creates a spectra fingerprint to compare against a database of known organisms

# MALDI-TOF

Analyte1



Analyte Name:



Analyte Description:

Analyte ID:



Analyte Creation Date/Time:



Applied MSP Library(ies):

Applied Taxonomy Tree:



Rank (Quality)	Matched Pattern	Score Value	NCBI Identifier
1 (++)	<i>Pseudomonas aeruginosa</i> ATCC 27853 THL	2.237	<a href="#">287</a>
2 (++)	<i>Pseudomonas aeruginosa</i> DSM 50071 T HAM	2.191	<a href="#">287</a>
3 (++)	<i>Pseudomonas aeruginosa</i> 8147_2 CHB	2.118	<a href="#">287</a>
4 (++)	<i>Pseudomonas aeruginosa</i> DSM 1117 DSM	2.108	<a href="#">287</a>
5 (+)	<i>Pseudomonas aeruginosa</i> 19955_1 CHB	1.908	<a href="#">287</a>
6 (+)	<i>Pseudomonas aeruginosa</i> A07_08_Pudu FLR	1.901	<a href="#">287</a>
7 (-)	<i>Pseudomonas jinjuensis</i> LMG 21316 T HAM	1.619	<a href="#">198616</a>
8 (-)	<i>Pseudomonas indica</i> DSM 14015 T HAM	1.437	<a href="#">137658</a>
9 (-)	<i>Pseudomonas citronellolis</i> DSM 50332 T HAM	1.388	<a href="#">53408</a>
10 (-)	<i>Pseudomonas taetrolens</i> LMG 2336 T HAM	1.346	<a href="#">47884</a>



# Version 1.0 Approved Organisms

<i>Acinetobacter baumanii</i>	<i>Micrococcus luteus</i>
<i>Aeromonas hydrophilia</i>	<i>Moraxella catarrhalis</i>
<i>Aggregatibacter aphrophilus</i>	<i>Neisseria gonorrhoeae</i>
<i>Arcanobacteria pyogenes</i>	<i>Neisseria lactamica</i>
<i>Bacteroides fragilis</i>	<i>Peptostreptococcus asaccharolyticus</i>
<i>Bacteroides uniformis</i>	<i>Prevotella melaninogenica</i>
<i>Burkholderia cepacia</i>	<i>Propionibacterium acnes</i>
<i>Campylobacter jejuni</i>	<i>Proteus mirabilis</i>
<i>Clostridium perfringens</i>	<i>Pseudomonas aeruginosa</i>
<i>Clostridium sordelli</i>	<i>Serratia marcescens</i>
<i>Corynebacterium diphtheriae</i>	<i>Staphylococcus aureus</i>
<i>Corynebacterium pseudodiphtheriticum</i>	<i>Staphylococcus epidermidis</i>
<i>Eikenella corrodens</i>	<i>Staphylococcus lugdunensis</i>
<i>Enterobacter aerogenes</i>	<i>Staphylococcus saprophyticus</i>
<i>Enterobacter cloacae</i>	<i>Stenotrophomonas maltophilia</i>
<i>Enterococcus faecalis</i>	<i>Streptococcus agalactiae</i>
<i>Enterococcus faecium</i>	<i>Streptococcus constellatus</i>
<i>Enterococcus gallinarum</i>	<i>Streptococcus dysgalactiae</i>
<i>Escherichia coli</i>	<i>Streptococcus equi</i>
<i>Fusobacterium necrophorum</i>	<i>Streptococcus pneumoniae</i>
<i>Haemophilus influenzae</i>	<i>Streptococcus pyogenes</i>
<i>Klebsiella oxytoca</i>	<i>Streptococcus sanguinis</i>
<i>Klebsiella pneumoniae</i>	

**Version 2.0 had hundreds. Version 3.0 at FDA now...**

# Performance of Matrix-Assisted Laser Desorption Ionization–Time of Flight Mass Spectrometry for Identification of Bacterial Strains Routinely Isolated in a Clinical Microbiology Laboratory<sup>▽</sup>

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Matrix-assisted laser desorption ionization–time of flight mass spectrometry (MALDI-TOF MS) has recently been introduced in diagnostic microbiology laboratories for the identification of bacterial and yeast strains isolated from clinical samples. In the present study, we prospectively compared MALDI-TOF MS to the conventional phenotypic method for the identification of routine isolates. Colonies were analyzed by MALDI-TOF MS either by direct deposition on the target plate or after a formic acid-acetonitrile extraction step if no valid result was initially obtained. Among 1,371 isolates identified by conventional methods, 1,278 (93.2%) were putatively identified to the species level by MALDI-TOF MS and 73 (5.3%) were identified to the genus level, but no reliable identification was obtained for 20 (1.5%). Among the 1,278 isolates identified to the species level by MALDI-TOF MS, 63 (4.9%) discordant results were initially identified. Most discordant results (42/63) were due to systematic database-related taxonomical differences, 14 were explained by poor discrimination of the MALDI-TOF MS spectra obtained, and 7 were due to errors in the initial conventional identification. An extraction step was required to obtain a valid MALDI-TOF MS identification for 25.6% of the 1,278 valid isolates. In conclusion, our results show that MALDI-TOF MS is a fast and reliable technique which has the potential to replace conventional phenotypic identification for most bacterial strains routinely isolated in clinical microbiology laboratories.

- Overall, ~93% success rate (2010 – databases continually improving)
- Generally:
  - Gram negative identification easier, more successful than Gram positives
  - More common organisms more successful than less common organisms
  - Certain bacteria (*Corynebacterium*, *Actinomycetes*), *Mycobacteria*, Molds, Yeasts lagging behind others, but catching up
  - Lack of identification (poor database coverage) much, much more likely than mis-identification
  - Exceptions in the case of several closely related species
    - NLF *E. coli* and *Shigella*
    - *Streptococcus pneumoniae* and *Streptococcus mitis*
    - Certain *Select Agents*

# Evaluation of Matrix-Assisted Laser Desorption/Ionization Time-of-Flight Mass Spectrometry for Identification of *Nocardia* species, and Other Aerobic Actinomyces

S. P. Buckwalter, S. L. Olson, B. J. Connelly, B. C. Lucas, A. A. Rodning, R. C. Walchak, Division of Clinical Microbiology, Department of Laboratory Medicine and Pathology, Mayo Clinic, Rochester, MN

The value of matrix-assisted laser desorption/ionization–time of flight mass spectrometry for identification of bacteria and yeasts is well documented in the literature. Its utility for the identification of *Nocardia* spp. has also been documented. We evaluated the performance of a matrix-assisted laser desorption/ionization–time of flight mass spectrometry-based identification system using both the performance of a reference report, our laboratory report, and a following library. The 16S rRNA gene sequence analysis of actinomycetes isolates is a valuable tool for the identification of *Nocardia* spp. and rapidly identify slow-growing

Matrix-Assisted Desorption Ionization Time-of-Flight Mass Spectrometry for the Use with Positive Blood Cultures: Methodology, Performance, and Optimization

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The use of matrix-assisted laser desorption/ionization time-of-flight mass spectrometry for smear-positive

Boaz Bishara, Neuberger

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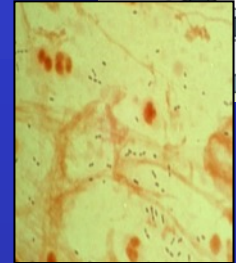
# Bacterial Culture and Identification Timeline



1 hr



1 hr

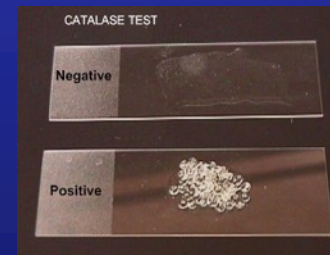
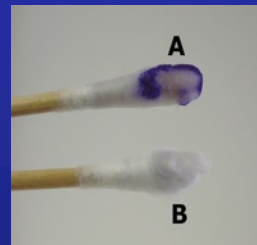


**Total time to ID: ~48 hours**  
**Total time to antibiotic profile: ~48 hours**

~18 hrs



~24 hrs



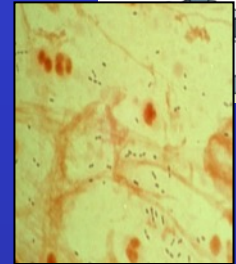
# Bacterial Culture and Identification Timeline



1 hr



1 hr



Total time to ID: ~24 hours

~18 hrs



10 min



~24 hrs





# Clinical Scenario #1

50 year old recently female retiree presents 1 year out from a right total knee replacement with worsening knee pain and mild swelling along with some night sweats. She is taken to the OR and the hardware is removed and sent down to the lab for culture... After 24 hours she has light growth of a short Gram negative rod growing only on the chocolate agar plate. MALDI identifies it as *Haemophilus influenzae*. She is treated with ceftriaxone for 6 weeks and her repeat cultures are negative and new hardware is placed and patient doing well to date.